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| (21) International Application Number: PCT/US92/09392 (22) International Filing Date: 2 November 1992 (02.11.92) (30) Priority data: 802,581 5 December 1991 (05.12.91) US (71) Applicant: PITMAN-MOORE, INC. [US/US]; 1401 S. Third Street, Terre Haute, IN 47808 (US). (72) Inventors: RAMAN, Siva, N. ; 815 Walfield Ct., St. Louis, MO 63141 (US). CUNNINGHAM, John, P. ; 1929 S. Fifth Street, Terre Haute, IN 47802 (US). (74) Agents: ERNST, Barbara, G. et al.; Rothwell, Figg, Ernst & Kurz, 555 13th Street, N.W. #701 East, Washington, DC 20004 (US). | | (81) Designated States: AU, BB, BG, BR, CA, CS, FI, HU, JP, KP, KR, LK, MG, MN, MW, NO, PL, RO, RU, SD, UA, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, SN, TD, TG). Published <i>With international search report.</i> |
| (54) Title: A CARBOHYDRATE GLASS MATRIX FOR THE SUSTAINED RELEASE OF A THERAPEUTIC AGENT (57) Abstract A composition for the sustained release of a biologically active therapeutic agent wherein the matrix of the sustained release composition is composed of an amorphous carbohydrate glass matrix comprising a suitable carbohydrate and an agent which retards the recrystallization of the carbohydrate and a biologically active therapeutic agent and a water-insoluble wax dispersed throughout the matrix. Biologically active therapeutic agents which can be incorporated into the carbohydrate glass matrix include prolactin, growth hormones, serum albumins, growth factors or any biologically active fragment or recombinant form thereof. | | |

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A CARBOHYDRATE GLASS MATRIX FOR THE
SUSTAINED RELEASE OF A THERAPEUTIC AGENT

Background of the Invention

The present invention is directed to a novel composition of matter for the sustained release of a therapeutic agent. More specifically, the present invention relates to a composition for the sustained release of a biologically active therapeutic agent from a biodegradable, amorphous carbohydrate glass matrix. Upon administration to a mammal, the amorphous carbohydrate glass matrix slowly dissolves, releasing the biologically active therapeutic agent into the physiological fluids of the animal.

The preparation and employment of amorphous carbohydrate glasses are known in the "candy industry" as well as in the production of medicated lozenges. However, amorphous carbohydrate glasses have not been employed as matrix material for sustained release polypeptide compositions. Various sustained release methods and compositions are known for administering therapeutic agents to both humans and animals alike. U.S. Patent No. 4,671,953, issued to Stanley et al. in 1987, describes a method and composition for administering sedatives, analgesics and anesthetics to a patient by incorporating the therapeutic agent into a lollipop. The lollipop is composed of a carbohydrate matrix or "candy matrix" with the therapeutic agent dispersed throughout the matrix. As the patient licks

or sucks on the "candy matrix" the matrix breaks down, releasing the drug into the oral cavity where the drug is absorbed across the mucosal cavity. A serious limitation to this method of administering a therapeutic agent to a patient is that the therapeutic agent must be sufficiently lipophilic that it will readily pass across a mucosal membrane. Examples of therapeutic agents which can be administered by this method include morphine, fentanyl, valium, midazolam and the like. Polypeptides are not sufficiently lipophilic such that they could be administered to a patient by this means.

U.S. Patent No. 4,765,980, issued to DePrince et al. in 1988, describes a method for stabilizing porcine growth hormone with porcine serum albumin in sustained release implant devices for swine. The porcine growth hormone and stabilizing amount of porcine serum albumin are compressed into a tablet or pellet with binders such as sodium bentonite, ethyl cellulose, stearic acid, adipic acid, fumaric acid, polyethylene glycol, deacetylated chitin and cellulose acetate. The pellets or tablets are loaded in a reservoir which can be implanted subcutaneously in swine where the growth hormone is slowly released from the reservoir. Typically, the matrix material of the reservoir is composed of polyalkylenes, polycarbonates, polyamides, modacrylic copolymers, polyesters and the like.

U.S. Patent No. 4,857,506, issued to Tyle in 1989, describes sustained release compositions of growth hormones in multiple water-in-oil-in-water emulsions. The compositions are administered subcutaneously to an animal where the growth hormone is slowly released from

the emulsion to increase the weight gain of the animal and increase milk production by a lactating animal.

U.S. Patent No. 4,293,539, issued to Ludwig et al. in 1981, describes controlled release formulations
5 useful in the prolonged treatment and control of microbial infections in animals. The antimicrobial agent is dispersed throughout a matrix composed of a copolymer of lactic acid and glycolic acid. The
10 copolymeric material is polymerized without a catalyst to ensure total degradation without leaving any toxic residues. The copolymer material then is dissolved in an organic solvent and mixed with a suitable
15 antimicrobial agent. The mixture is extruded into a desired shape and then cooled to form a hard glass-like device. Antimicrobials typically employed include penicillins, cephalosporins, tetracyclines, sulfa
drugs, macrolide antibiotics and aminoglycosides.

U.S. Patent No. 2,918,411, issued to Hill in 1959, describes a controlled release oral pharmaceutical
20 dosage form. The essential components of the formulation include polyvinylpyrrolidone, a substantially water-insoluble agent, such as a saturated fatty acid, a saturated fatty acid ester and a pharmacologically acceptable sterol, and a
25 pharmacologically active material. After the essential components of the formulation are mixed, the formulation is pelletized. The pellets then can be encapsulated or embodied in another type of dosage form
or administered directly to a patient from the bulk
30 pellet. Suitable pharmacologically active agents which can be employed in this sustained release composition include antibiotics, hypnotics, tranquilizing agents, antihistamines and narcotics.

Although there is a variety of sustained release compositions for many biologically active agents, there is still a need for a sustained release composition which completely degrades in the physiological fluids of the host leaving behind little, if any, residual material and which does not substantially interact with the therapeutic agent.

Summary of the Invention

The present invention is directed to a sustained release composition for biologically active therapeutic agents comprising a biodegradable, amorphous carbohydrate glass matrix, a hydrophobic substance, and a biologically active therapeutic agent dispersed throughout the matrix. The composition can be administered orally to or implanted in an animal where the amorphous carbohydrate glass matrix slowly degrades, releasing a biologically active therapeutic agent into the physiological fluids of the animal.

Detailed Description of the Invention

The present invention relates to a novel sustained release composition comprising a biologically active therapeutic agent dispersed throughout an amorphous carbohydrate glass matrix. More specifically, sustained release compositions of the present invention comprise a biodegradable, amorphous carbohydrate glass matrix, an agent to retard crystallization of the amorphous carbohydrate matrix, a biologically active therapeutic agent and a hydrophobic substance to modify the rate of release of the polypeptide from the amorphous carbohydrate matrix.

The composition of the present invention can be administered orally or by implanting subcutaneously

such that the amorphous carbohydrate matrix slowly dissolves, releasing the therapeutic agent into the physiological fluids wherein the therapeutic agent asserts its biological activity on the animal.

5 Advantageously, the amorphous carbohydrate matrix does not substantially interact with therapeutic agents such that the biological activity of a therapeutic agent incorporated within the matrix would be compromised. The amorphous carbohydrate glass matrix completely
10 degrades in the physiological fluids of the host leaving very little, if any, residual material behind to be removed after release of the therapeutic agent is complete. Also advantageously, the composition has a relatively slow rate of dissolution in physiological
15 fluids, dissolution occurs predominantly at the surface of the carbohydrate matrix, and the sustained release compositions within the scope of this invention are amenable to conventional production techniques such as extrusion, tableting and the like.

20 Generally, the amorphous carbohydrate component, including the recrystallization retarding agent, comprises from about 60% to about 90% by weight of the sustained release composition. The amorphous carbohydrate component comprises one or more
25 carbohydrates mixed with an agent to retard recrystallization of the carbohydrate. Suitable carbohydrates which can be employed as matrix material include, but are not limited to, disaccharides such as sucrose, lactose, maltose or cellobiose. Suitable
30 agents employed to retard recrystallization of the carbohydrate include, but are not limited to, polyvinylpyrrolidone (PVP), polyethylene glycols, polyvinyl alcohol, maltodextrins, sodium lauryl sulfate, oleyl alcohol, stearyl alcohol and the like.

Generally, the carbohydrate component comprises from about 50% to about 80% by weight of the amorphous carbohydrate component and the recrystallization retarding agent comprises from about 10% to about 40% by weight of the amorphous carbohydrate component. Water comprises the rest of the amorphous carbohydrate component.

Therapeutic agents which can be employed in the sustained release compositions of the present invention include biologically active polypeptides, antibiotics and vitamins. As used herein, the term "polypeptide" encompasses natural and recombinant polypeptides having a desired biological activity, including bioactive polypeptides having deleted, replaced or altered amino acid sequences in comparison with the full-length natural polypeptide.

Polypeptides which can be incorporated within the amorphous carbohydrate glass matrices generally have a molecular weight of at least 1000 daltons and no more than about 200,000 daltons. The polypeptide component of the sustained release compositions of the present invention ranges from about 2% to about 20% by weight of the composition. Examples of polypeptides which can be employed to practice this invention include, but are not limited to, prolactin (PRL), serum albumin, growth factors and growth hormones, i.e., somatotropin. Serum albumins which can be employed to practice this invention include but are not limited to, bovine, ovine, equine, avian and human serum albumin. Examples of suitable growth factors include epidermal growth factor (EGF), insulin-like growth factor I (IGF-I), insulin-like growth factor II (IGF-II), fibroblast growth factor (FGF), transforming growth factor alpha (TGF- α), transforming growth factor beta (TGF- β), nerve

growth factor (NGF), and platelet-derived growth factor (PDGF). Particularly preferred growth factors include recombinant human insulin-like growth factors I (rHuIGF-I) and II (rHuIGF-II). Somatotropins which can be employed to practice this invention include, but are not limited to, bovine, porcine, ovine, equine, avian and human somatotropin. A preferred porcine somatotropin is delta-7 recombinant porcine somatotropin, described and claimed in European Patent Application Publication No. 104,920 (Biogen).

Hydrophobic substances which can be employed to modify the rate of release of a therapeutic agent from an amorphous carbohydrate glass matrix include water insoluble waxes such as white beeswax, yellow beeswax, candelilla, carnauba wax, castor wax, cetyl esters wax and the like. Other materials such as cholesterol glycerol monostearate, fatty acid esters of glycerol, fatty acids such as stearic acid, fats, lipids, etc. also can be employed. Generally, these water-insoluble substances comprise from 5% to about 25% by weight of the sustained release composition.

Optionally, the formulation also may contain aciduants, preservatives, etc. known to those skilled in the art.

The sustained release compositions of the present invention can be prepared by a variety of methods. Examples of suitable methods for preparing amorphous carbohydrate glass matrices are disclosed in Pharmaceutical Dosage Forms, Vol. 1, chapter 8, "Medicated Lozenges" by D. Peters (H. Lieberman and L. Lachman, eds.) herein incorporated by reference. A preferred method comprises mixing an aqueous solution of a carbohydrate, wherein the carbohydrate ranges from about 60% to about 80% by weight of the solution, with

an aqueous solution of a recrystallization retarding agent. Generally, a recrystallization retarding agent comprises from about 25% to about 40% by weight of the solution. The two aqueous solutions are thoroughly
5 mixed and heated to a temperature ranging from about 110°C to about 120°C until the mixture becomes viscous. The viscous mixture then is allowed to cool to a temperature of from about 60° to about 70°C. A suitable biologically active therapeutic agent then is
10 thoroughly kneaded into the viscous carbohydrate composition using any suitable technique, such as by hand. A water insoluble substance as described above also can be added and kneaded into the viscous carbohydrate-therapeutic agent mixture.

15 The resulting mixture can be extruded through preheated syringes at temperatures ranging from about 60° to about 70°C to form rods. The viscous carbohydrate-therapeutic agent mixture also can be shaped into tablets and the like according to methods practiced in
20 the art. The shaped carbohydrate-therapeutic agent mixture is allowed to cool to room temperature (i.e., about 18-25°C), thus forming a sustained release composition comprising an amorphous carbohydrate glass matrix having a biologically active therapeutic agent
25 dispersed throughout the matrix. Sustained release compositions within the scope of this invention can be implanted subcutaneously in an animal where the amorphous carbohydrate glass matrix slowly erodes, releasing a biologically active therapeutic agent into
30 the physiological fluids of the host. Other modes of administration, such as oral dosing, also can be employed.

The following examples are intended to particularly point out and more clearly illustrate the invention and are not intended to be limiting.

Example 1

5 An aqueous sucrose solution (70% by weight, 24.11 g) was mixed with an aqueous solution of polyvinylpyrrolidone (PVP) (33% by weight, 34.10 g) and heated to about 115-118°C. As the water in the mixture continued to evaporate, the mixture became viscous and
10 turned pale yellow to deep amber in color. The hot viscous mixture was poured into a beaker and cooled to room temperature (about 20°C) yielding a solid amorphous sucrose matrix (ASM). The X-ray diffraction pattern, determined using a Philips 3100 X-ray Powder
15 Diffractometer, indicated an amorphous structure. The water content of the ASM was about 11.8% by weight and the reducing sugar content was about 1.4%. The water content was determined using the commonly known Karl Fischer Moisture Determination method. The amount of
20 reducing sugar was determined by the reduction of alkaline ferricyanide and subsequent calorimetric analysis at 410 nm.

Example 2

25 ASM (1.5 g) as prepared in Example 1 was placed on a Teflon® sheet preheated to about 60° to 70°C and thoroughly kneaded with 0.15 g of solid particles of bovine serum albumin (BSA) (obtained from Sigma Chemical Co.) Molten beeswax (0.302 g) was added to the BSA-ASM mixture and kneaded to give Formulation A
30 comprising: ASM (77 parts), BSA (8 parts) and beeswax (15 parts). The resulting viscous formulation was heated to about 70°C and loaded into pre-heated plastic

syringes also at about 70°C. The formulation was extruded to give cylindrical rods. Short sections of about 5 mm in length were cut from these rods to give 6 cylindrical pieces weighing 110-135 mg. Each cylindrical rod was placed in a culture tube containing 5 ml of 10mM phosphate buffered saline (PBS) at a pH of 7.4. The cylindrical pieces were shaken in a Gyrotory® water-bath at 37°C. Periodically, the solutions were removed and replaced with fresh PBS. BSA concentrations in the buffer solutions were determined by measuring the UV absorbances at 278 nm with a Beckman spectrophotometer (Model DU-70). Release profiles were constructed by calculating the cumulative percent of BSA released at various time intervals. Table 1 discloses the mean values of the cumulative percent of BSA released at different time periods.

TABLE 1
Release of BSA from Formulation A

| Time | Cumulative % of BSA Released | |
|---------|------------------------------|--------------------|
| | Mean | Standard Deviation |
| 20 Days | | |
| 0.083 | 22.0 | 4.6 |
| 0.33 | 32.7 | 8.0 |
| 1.0 | 45.5 | 10.9 |
| 2.0 | 55.4 | 11.8 |
| 25 3.0 | 61.9 | 12.0 |
| 4.0 | 66.6 | 11.8 |
| 5.0 | 70.6 | 11.6 |

The pattern of release of BSA shows that an amorphous carbohydrate glass matrix can be employed for the sustained release of a polypeptide.

Examples 3 to 5

Formulations B, C and D were prepared according to the methods described in Examples 1 and 2 above except that the amount of beeswax in each formulation differed. Formulation A in Example 2 comprised 15 parts by weight of beeswax while Formulations B, C and D comprised 8.3, 4.4 and 0 parts by weight of beeswax, respectively. The release rates were determined by the same method as in Example 2. The compositions of Formulations B, C and D were as follows:

Formulation B

| | <u>Component</u> | <u>Amount</u> |
|----|------------------|---------------------|
| | ASM | 1.50 g (83.3 parts) |
| | BSA | 0.15 g (8.3 parts) |
| 15 | Beeswax | 0.15 g (8.3 parts) |

The release of BSA from the cylinders was complete in about 3.5 hours.

Formulation C

| | <u>Component</u> | <u>Amount</u> |
|----|------------------|---------------------|
| 20 | ASM | 1.50 g (87 parts) |
| | BSA | 0.15 g (8.7 parts) |
| | Beeswax | 0.075 g (4.4 parts) |

The release of BSA from the cylinders was complete in about 1.75 hours.

Formulation D

| | <u>Component</u> | <u>Amount</u> |
|----|------------------|-------------------|
| 25 | ASM | 1.50 g (91 parts) |
| | BSA | 0.150 g (9 parts) |
| | Beeswax | 0.00 g |

30 The release of BSA from the cylindrical pieces was complete in about 1 hour.

The results from this experiment indicate that the release rate of a polypeptide from an amorphous carbohydrate glass matrix can be modified by varying the amount of wax in the sustained release composition. The higher the wax content in the formulation, the slower was the release of BSA.

Examples 6 and 7

Formulation E and F were prepared according to the methods of Examples 1 and 2 except that a zinc complex of recombinant porcine somatotropin (ZnnpST) (prepared by Pitman-Moore, lot. # 148/010/011-820) replaced the BSA as the polypeptide in the sustained release composition. The formulation of composition E was as follows:

| <u>Formulation E</u> | | |
|----------------------|---------------|--|
| <u>Component</u> | <u>Amount</u> | |
| ZnnpST | 0.15 g | |
| ASM | 1.50 g | |
| Beeswax | 0.30 g | |

TABLE 2
Release of ZnnpST from Formulation E

| <u>Time</u> <u>Days</u> | <u>Cumulative % of ZnnpST Released</u> | |
|----------------------------|--|---------------------------|
| | <u>Mean</u> | <u>Standard Deviation</u> |
| 0.33 | 16.8 | 3.9 |
| 1.0 | 22.2 | 4.8 |
| 2.0 | 25.2 | 4.4 |
| 3.0 | 27.1 | 3.8 |

Formulation F

| <u>Component</u> | <u>Amount</u> |
|------------------|---------------|
| ASM | 1.50 g |
| ZnnpST | 0.15 g |

5 Release of ZnnpST from Formulation F was complete in
about 1 hour. The beeswax incorporated into
Formulation E clearly reduced the rate of release of
ZnnpST from the carbohydrate matrix, since by the third
10 day only 27.1% of the ZnnpST had been released. This
experiment clearly showed that a carbohydrate glass
matrix can be employed for the sustained release of a
somatotropin.

In order to find out whether the formulation
techniques had affected the biological activity of the
15 pST, radioreceptor binding assays were performed on
samples of porcine somatotropin (pST) extracted from
Formulation E. PST extracted from Formulation E had
about 87% of the binding activity of the protein prior
to formulation. The results indicated that the pST
20 retained its activity substantially in the course of
the formulation steps.

We claim:

1. A sustained release composition comprising an amorphous carbohydrate glass matrix, a biologically active therapeutic agent, and a hydrophobic substance which modifies the rate of release of the therapeutic agent from the glass matrix.

2. A sustained release composition according to claim 1, wherein the amorphous carbohydrate glass matrix comprises a carbohydrate and an agent which retards recrystallization of the amorphous carbohydrate glass matrix.

3. A sustained release composition according to claim 2, wherein the amorphous carbohydrate matrix comprises from about 50%-75% by weight of the matrix and the recrystallization retarding agent comprises from about 15%-40% by weight of the matrix.

4. A sustained release composition according to claim 1, wherein the hydrophobic substance is a wax.

5. A sustained release composition according to claim 4, wherein the wax comprises white beeswax, yellow beeswax, candelilla wax, carnauba wax, vegetable waxes, castor wax or cetyl esters wax.

6. A sustained release composition according to claim 1, wherein the hydrophobic substance comprises cholesterol, fatty acid esters or fatty acids.

7. A sustained release composition according to claim 1, wherein the hydrophobic substance comprises from about 5% to about 25% by weight of the composition.

8. A sustained release composition according to claim 2, wherein the carbohydrate comprises sucrose, lactose, maltose or cellobiose.

9. A sustained release composition according to claim 1, wherein the amorphous carbohydrate matrix

comprises from about 60% by weight to about 90% by weight of the sustained release composition.

10. A sustained release composition according to claim 1, wherein the therapeutic agent comprises a polypeptide, vitamin or antibiotic.

11. A sustained release composition according to claim 1, wherein the polypeptide comprises prolactin, serum albumins, somatotropins, growth factors or any biologically active fragment or recombinant form thereof.

12. A sustained release composition according to claim 11, wherein the therapeutic agent comprises from about 2% to about 20% by weight of the composition.

13. A sustained release composition of claim 11, wherein the serum albumin comprises bovine, ovine, porcine, avian or human serum albumin.

14. A sustained release composition according to claim 11, wherein the somatotropin comprises bovine, ovine, porcine, avian or human somatotropin.

15. A sustained release composition according to claim 11, wherein the growth factor comprises epidermal growth factor, insulin-like growth factor I, insulin-like growth factor II, fibroblast growth factor, transforming growth factor alpha, transforming growth factor beta, platelet-derived growth factor or nerve growth factor.

16. A sustained release composition according to claim 2, wherein the recrystallization retarding agent comprises polyvinylpyrrolidone, polyvinyl alcohol, polyethylene glycols, maltodextrins, sodium lauryl sulfate, oleyl alcohol or stearyl alcohol.

17. A sustained release composition according to claim 1, wherein the amorphous carbohydrate glass matrix comprises from about 60%-90% by weight of the

composition; the biologically active therapeutic agent comprises from about 2%-20% by weight of the composition; and the wax comprises from about 5% to about 25% by weight of the composition.

5 18. A sustained release composition comprising a mixture of recombinant porcine somatotropin; polyvinylpyrrolidone, and beeswax dispersed in an amorphous sucrose glass matrix.

10 19. A method for preparing a sustained release composition comprising:

- a. Mixing an aqueous sucrose solution with an aqueous solution of polyvinylpyrrolidone;
- b. heating the mixture at a temperature of from about 115°C to about 118°C to evaporate a sufficient
15 amount of water such that the mixture becomes viscous;
- c. cooling the viscous mixture to a temperature of from about 60°C to about 70°C;
- d. kneading a zinc complex of recombinant porcine somatotropin and beeswax with the viscous
20 mixture of sucrose and polyvinylpyrrolidone;
- e. extruding the viscous mixture of sucrose, polyvinylpyrrolidone, beeswax and somatotropin to form rods;
- f. cutting the rods to form short cylindrical
25 sections; and
- g. cooling the short cylindrical sections to form sustained release cylinders having an amorphous sucrose and polyvinylpyrrolidone glass matrix and somatotropin and beeswax dispersed throughout the
30 matrix.

20. A method of administering a biologically active therapeutic agent to a mammal comprising inserting a sustained release composition subcutaneously in said mammal wherein the composition

comprises an amorphous carbohydrate glass matrix having
a biologically active therapeutic agent and water-
insoluble wax incorporated within the matrix whereby
the carbohydrate glass matrix is degraded by the
5 physiological fluids of the animal, releasing the
polypeptide from the matrix.

21. A method of administering a biologically
active therapeutic agent to a mammal by administering
orally a sustained release composition wherein the
10 composition comprises an amorphous carbohydrate glass
matrix having a biologically active therapeutic agent
and water-insoluble wax incorporated within the matrix
whereby the carbohydrate glass matrix is degraded by
the physiological fluids of the animal, releasing the
15 polypeptide from the matrix.

INTERNATIONAL SEARCH REPORT

PCT/US 92/09392

International Application No

I. CLASSIFICATION F SUBJECT MATTER (if several classification symbols apply, indicate all)⁶

According to International Patent Classification (IPC) or to both National Classification and IPC

Int.Cl. 5 A61K9/00; A61K47/26; A61K47/44

II. FIELDS SEARCHEDMinimum Documentation Searched⁷

| Classification System | Classification Symbols |
|-----------------------|------------------------|
| Int.Cl. 5 | A61K |

Documentation Searched other than Minimum Documentation
to the Extent that such Documents are Included in the Fields Searched⁸**III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹**

| Category ¹⁰ | Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹² | Relevant to Claim No. ¹³ |
|------------------------|--|-------------------------------------|
| Y | WO,A,8 904 689 (BAXTER INTERNATIONAL) 1 June 1989 see claims 1-5,9-11,33,39-42 see page 5, line 20 - line 22 see page 6, line 9 - line 16 see page 8, line 19 - line 26 see page 11, line 7 - line 21 see page 16, line 6 - line 13 see page 18, line 11 - line 16 | 1,3-4,6, 8-10,12, 17,19 |
| Y | EP,A,0 345 628 (ABBOTT) 13 December 1989 see claims see page 3, line 31 - line 56 see page 4, line 24 - line 30 -/- | 1,3-4,6, 8-10,12, 17,19 |

¹⁰ Special categories of cited documents : ¹⁰¹⁰ "A" document defining the general state of the art which is not considered to be of particular relevance¹⁰ "E" earlier document but published on or after the international filing date¹⁰ "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)¹⁰ "O" document referring to an oral disclosure, use, exhibition or other means¹⁰ "P" document published prior to the international filing date but later than the priority date claimed¹⁰ "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention¹⁰ "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step¹⁰ "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.¹⁰ "A" document member of the same patent family**IV. CERTIFICATION**

Date of the Actual Completion of the International Search

22 FEBRUARY 1993

Date of Mailing of this International Search Report

04.03.93

International Searching Authority

EUROPEAN PATENT OFFICE

Signature of Authorized Officer

SCARPONI U.

| III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET) | | Relevant to Claim No. |
|--|---|-------------------------------|
| Category ^a | Citation of Document, with indication, where appropriate, of the relevant passages | |
| Y | FR,A,2 383 659 (BERRI-BALZAC) 13 October 1978 see claims see page 1, line 9 - line 10 see page 2, line 14 - line 18 see page 4, line 3 - line 6 see example 2 | 1,3-4,6, 8-10,12, 17,19 |
| Y | US,A,2 918 411 (J.A.HILL) 22 December 1959 cited in the application See the whole document | 1,3-4,6, 8-10,12, 17,19 |
| Y | WO,A,9 103 237 (UNIVERSITY OF UTAH) 21 March 1991 cited in the application see claims 1-8,13,17-19,21-23 see page 9, line 20 - line 24 see page 10, line 13 - line 22 see page 12, line 15 - line 25 see page 40, line 7 - line 11 | 1,3-4,6, 8-10,12, 17,19 |

**ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO.**

US 9209392
SA 67217

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report.
The members are as contained in the European Patent Office EDP file on
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For more details about this annex : see Official Journal of the European Patent Office, No. 12/82